

Hands-on QM/MM Tutorial

Files are also available on:

<http://villekaila.wordpress.com/>

and on the CHARMM web page

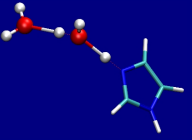
Our files are also provided on

<http://github.com/RowleyGroup/charmm-turbomole-examples/tree/master/qmmm>

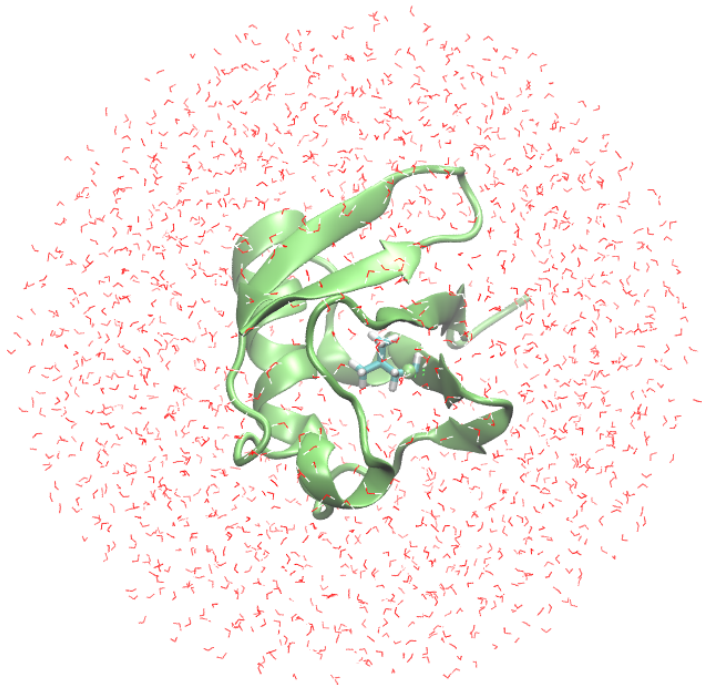
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Prof. Ville R. I. Kaila
16.3.2018



■ Example: QM/MM MD simulation of ubiquitin in water:



Build system using

1) **VMD**

Extensions → Modeling → AutoPSF
→ Add Solvation → Add Ions
(not recommended for complex systems)

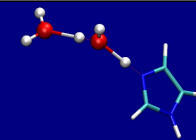
or write psfgen script
example make.tcl
followed by **solvate** and **autoionize**
(recommend for complex systems)

or

2) **CHARMM-GUI**

a nice web-based portal, which also
gives you a charmm input file

Solvating the protein with VMD



see NAMD/VMD tutorials

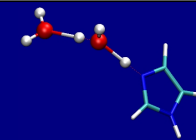
<http://www.ks.uiuc.edu/Training/Tutorials/namd-index.html>

VMD has the commands:

```
solvate protein.psf protein.pdb -t 15 -o protein-wb
```

```
autoionize -psf protein-wb.psf -pdb protein-wb.pdb -sc 0.15
```

Using CHARMM-GUI to prepare input files





CHARMM-GUI

Effective Simulation Input Generator and More

CHARMM is a versatile program for atomic-level simulation of many-particle systems, particularly macromolecules of biological interest. - M. Karplus

[about us](#) :: [input generator](#) :: [archive](#) :: [charmm docs](#) :: [lectures](#) :: [movie gallery](#) :: [video demo](#) :: [citations](#) :: [update log](#) :: [jobs & events](#) :: [giving](#)

Some [lectures](#) and [job postings](#) are now available. See [upload log](#) for update history and [giving](#) for donation. [Contact](#) info is given below.

CHARMM-GUI

- About Us
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- ST-analyzer
- Archive
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Front Page

CHARMM-GUI provides a web-based graphical user interface to generate various molecular simulation systems and input files to facilitate and standardize the usage of common and advanced simulation techniques. Currently, CHARMM-GUI supports CHARMM, NAMD, GROMACS, AMBER, GENESIS, LAMMPS, Desmond, OpenMM, and CHARMM/OpenMM simulation programs mostly based on the CHARMM force fields.

CHARMM-GUI is powered by CHARMM, an academic research program used world-wide for macromolecular dynamics and mechanics (<http://www.charmm.org>). Its development began in the research group of Professor Martin Karplus at Harvard University and continues throughout the world with contributing developers. CHARMM performs standard molecular dynamics and energy minimization with the potential energy functions for proteins, nucleic acids, lipids, carbohydrates, and various small molecules. In addition, CHARMM can be used for various chemical and conformational free energy calculations with many types of restraints.

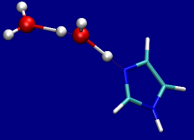
The CHARMM-GUI team hopes that the tools and materials offered here are useful and helpful for your research and education.

Academic users can obtain CHARMM freely from <http://www.charmm.org>.

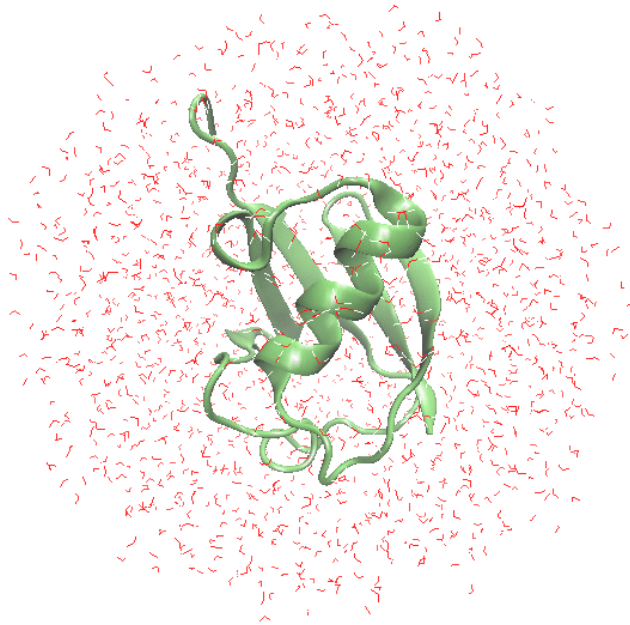
Geographical Visitors



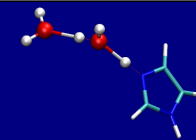
MD relaxation



- Before you start with QM/MM simulations → properly relax the system by classical MD



QM/MM input for CHARMM



```
* QM/MM calculation of a protein, water, ion system  
*
```

Charmm files start with
dot -- dot

```
! QM/MM protein tutorial  
! by Ville R. I. Kaila  
! Technische Universitaet Muenchen  
! Department Chemie  
! E-mail: ville.kaila@ch.tum.de  
! Ubiquitin in water
```

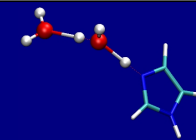
This is a comment

```
! qturboexe points to the "turbo.py" script that executes TURBOMOLE
```

```
envi qturboexe "/wrk/vkaila/DONOTREMOVE/SPRING-  
SCHOOL/SPRINGSCHOOL/qmmm_protein/charmm_input/turbo711.py"
```

set variable
turbo711.py
python code for
QM/MM

QM/MM input for CHARMM



```
envi qturbooutpath "turbodir"
```

```
! "qturboinpath" includes the input files for  
TURBOMOLE, e.g. control, basis, ...  
envi qturboinpath "data"
```

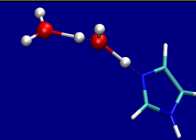
```
! creating the directories in the current working  
directory  
system "mkdir -p turbodir"  
system "mkdir -p turbo_tmp"  
system "mkdir -p data"  
system "mkdir -p charmm_output"
```

"qturbooutpath" is the directory that includes "coord", "point_charges", and "gradient" files that are mutual between CHARMM and TURBOMOLE

(coord and point charges are updated ! before every QM calculation by CHARMM, gradient is provided by TURBOMOLE and read by CHARMM)

create
directories where
the TURBOMOLE
files are run

QM/MM input for CHARMM



```
BOMBLEV -2
```

```
set a "toppar/top_all22_prot.rtf"  
set b "toppar/par_all22_prot.prm"  
set c "toppar/toppar_water_ions.str"
```

```
! read in a, b, c  
read rtf card name @a
```

```
read para card flex name @b
```

```
! append the nucleic acid parameters  
read para card flex append name @c
```

```
OPEN UNIT 1 READ CARD NAME "toppar/qqh.rtf"  
READ RTF CARD UNIT 1 append  
CLOSE UNIT 1
```

```
OPEN UNIT 1 READ CARD NAME "toppar/qqh.prm"  
READ PARA CARD UNIT 1 append flex  
CLOSE UNIT 1
```

sets debugging level in CHARMM
goes to -5 (only for testing stuff!)

read in topology & parameter files for
MM

a bit charmming by creating variables
a, b, c and then reading them

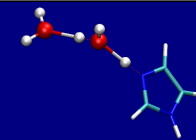
the flex command is used for appending

here we read in parameters for the
link atoms
(we need dummy parameters with
"zeros")

topology file starts
with dot dots

and then mass of all elements

Lets have a look at the CHARMM topology file

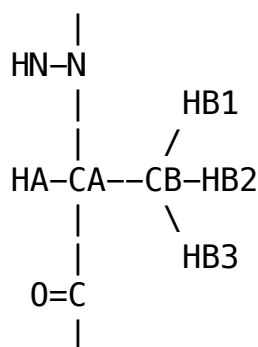


```
RESI ALA 0.00
GROUP
ATOM N NH1 -0.47 !
ATOM HN H 0.31 ! HN-N
ATOM CA CT1 0.07 !
ATOM HA HB 0.09 ! HB1
GROUP !
ATOM CB CT3 -0.27 ! HA-CA--CB-HB2
ATOM HB1 HA 0.09 ! HB3
ATOM HB2 HA 0.09 !
ATOM HB3 HA 0.09 !
GROUP !
ATOM C C 0.51
ATOM O O -0.51
```

atom name
(as in **pdb** file)

atom type
(refers to the parameter file)
atom types & charges
are written to the **psf** file

resname total charge/partial charges



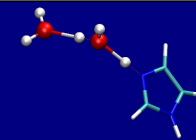
create bond between CB and CA
create bond between N HN
etc.

CHARMM
backbone correction CMAP

```
BOND CB CA N HN N CA
BOND C CA C +N CA HA CB HB1 CB HB2 CB HB3
DOUBLE O C
IMPR N -C CA HN C CA +N O
CMAP -C N CA C N CA C +N
DONOR HN N
ACCEPTOR O C
IC -C CA *N HN 1.3551 126.4900 180.0000 115.4200 0.999
IC -C N CA C 1.3551 126.4900 180.0000 114.4400 1.539
....
```

internal coordinate list
(helps to build the residue)

Lets have a look at the CHARMM parameter file (1/3)



BONDS

$V(\text{bond}) = K_b(b - b_0)^2$

!Kb: kcal/mole/A**2

!b0: A

!atom type Kb b0

CA CA 305.000 1.3750

...

force constant and eq. distance
for bonding term between atom
types CA and CA

ANGLES

$V(\text{angle}) = K_{\theta}(\theta - \theta_0)^2$

!Ktheta: kcal/mole/rad**2

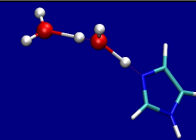
!Theta0: degrees

CT1 CT1 C 52.000 108.0000

...

force constant and eq. distance
for angle terms
between atom types CT1-CT1-C

Lets have a look at the CHARMM parameter file (2/3)



DIHEDRALS

```
!  
!V(dihedral) = Kchi(1 + cos(n(chi) - delta))  
!
```

```
!Kchi: kcal/mole  
!n: multiplicity  
!delta: degrees  
!
```

```
!atom types          Kchi    n    delta  
!  
CE2  CE1  CT2  HA      0.1200  3      0.00 !
```

force constant and eq. dihedrals
for dihedral terms
between atom types
CE2-CE1-CT2-HA

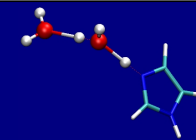
IMPROPER

```
!  
!V(improper) = Kpsi(psi - psi0)**2  
!  
!Kpsi: kcal/mole/rad**2  
!psi0: degrees  
!note that the second column of numbers (0) is ignored  
!
```

```
!atom types          Kpsi          psi0  
0      X      X      C      120.0000      0      0.0000
```

atom types
O-X-X-C kept at 120 deg
planarity

Lets have a look at the CHARMM parameter file (3/3)

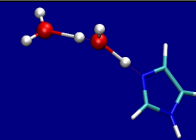


```
!V(Lennard-Jones) = Eps,i,j[(Rmin,i,j/ri,j)**12 - 2(Rmin,i,j/ri,j)**6]
!  
!epsilon: kcal/mole, Eps,i,j = sqrt(eps,i * eps,j)  
!Rmin/2: A, Rmin,i,j = Rmin/2,i + Rmin/2,j  
!  
!atom  ignored      epsilon      Rmin/2  ignored  eps,1-4      Rmin/2,1-4  
SOD      0.0          -0.0469      1.41075
```

Lennard-Jones terms for
atom types "SOD"

... lets now continue with input file

QM/MM input contd.



```
open read card unit 10 name "segments/test_proa.pdb"
read sequence pdb unit 10
generate PROA setup warn first NTER last CTER
```

read in protein segment PROA
create NTER and CTER patch
(look at PRES NTER/CTER in
topology file)

```
read coor pdb unit 10 resid
```

```
! read in water segment WATA
open read card unit 10 name "segments/test_wata.pdb"
read sequence pdb unit 10
generate WATA setup warn noangle nodihedral
```

read in water segment WATA
.. in charmm you need to have
different segments for protein
subunits, water, ions, etc.

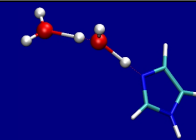
```
read coor pdb unit 10 resid
```

YES IT IS LIKE FORTRAN...

```
!open read card unit 10 name "./sod.pdb"
!read sequence pdb unit 10
!generate I01 setup warn noangle nodihedral
```

read in ions
→ not in this example
(we commented them out)

QM/MM input contd.



```
! print out pdb structure
open write card unit 10 name "charmm_output/test_all.pdb"
write coor pdb unit 10
```

write combined
coordinates into
pdb , crd,

```
! start from the coordinates of
! a previous run
!open read unit 22 card name "charmm_input/mini_all.crd"
!read coor unit 22 card
```

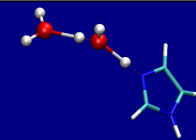
```
!OPEN UNIT 31 READ CARD
NAME "charmm_input/structure.psf"
!READ PSF CARD UNIT 31
!CLOSE UNIT 31
```

if you already have a crd and psf
file you can read in them instead
of creating your system

```
!OPEN UNIT 31 READ CARD
NAME "charmm_input/structure.crd"
!READ COOR CARD UNIT 31
!CLOSE UNIT 31
```

NOTE: charmm has a different
psf format than NAMD, which
uses xplor format

QM/MM input contd.



ENERGY

... now finally starts QM/MM specific part

```
addlinkatom QQH1 PROA 43 CB PROA 43 CA
               order qm  - mm
```

classical single point energy

create a link atom "QQH1"
between CB and CA of resid 43
in segname PROA

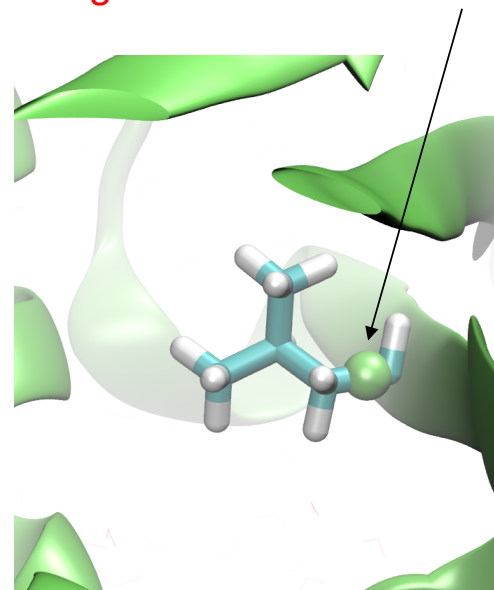
```
define qm sele -
atom PROA 43 CB -
.or. atom PROA 43 HB1 -
.or. atom PROA 43 HB2 -
.or. atom PROA 43 CG -
.or. atom PROA 43 HG -
.or. atom PROA 43 CD1 -
.or. atom PROA 43 HD11 -
.or. atom PROA 43 HD12 -
.or. atom PROA 43 HD13 -
.or. atom PROA 43 CD2 -
.or. atom PROA 43 HD21 -
.or. atom PROA 43 HD22 -
.or. atom PROA 43 HD23 -
show end
```

line continues:

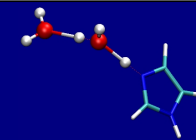
define the qm system

NOTE:

There is no physical/chemical purpose
of simulating resid 43 by QM/MM
(example is only for demonstration purposes)



QM/MM input contd.



```
lonepair colinear scaled -.7261 -  
sele type QQH1 show end sele atom PROA 43 CB -  
show end sele atom PROA 43 CA show end
```

```
print coord sele qm .or. atom * * QQH* end
```

```
stop
```

execute:

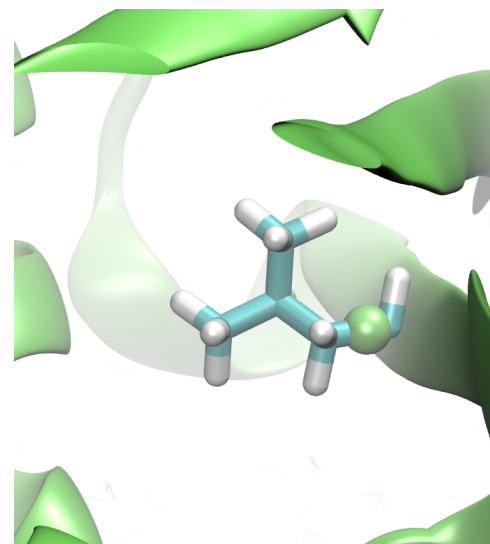
```
./charm  < qmmm_protein.inp > qmmm_protein.out
```

print the qm region and stop the code

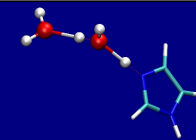
→

take coordinates and perform TURBOMOLE single point

in order to place the link atom in
the correct direction use lone
pair command



TURBOMOLE single point for QM/MM (1/5)



cd data

prepare coord.xyz based on what charmm prints out that contains the qm region

x2t coord.xyz > coord for this use need: module load turbomole

define

IF YOU WANT TO READ DEFAULT-DATA FROM ANOTHER control-TYPE FILE,
THEN ENTER ITS LOCATION/NAME OR OTHERWISE HIT >return<.

hit enter

INPUT TITLE OR

how to run define

ENTER & TO REPEAT DEFINITION OF DEFAULT INPUT FILE

hit enter

IF YOU APPEND A QUESTION MARK TO ANY COMMAND AN EXPLANATION
OF THAT COMMAND MAY BE GIVEN

a coord

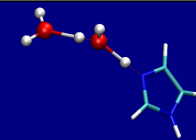
IF YOU APPEND A QUESTION MARK TO ANY COMMAND AN EXPLANATION
OF THAT COMMAND MAY BE GIVEN

*

IF YOU DO NOT WANT TO USE INTERNAL COORDINATES ENTER no

no ! internal coordinates will mess up the qm system positioning
relative to mm point charges

TURBOMOLE single point for QM/MM (2/5)



ATOMIC ATTRIBUTE DEFINITION MENU (#atoms=198 #bas=198 #ecp=2)

b

ENTER A SET OF ATOMS TO WHICH YOU WANT TO ASSIGN BASIS SETS

(ATOMIC SET : all none)

TO OUTPUT ATOMIC SET SYNTAX ENTER A QUESTION MARK ?

E.G. all dz (DZ BASIS FOR ALL ATOMS)

all sto-3g hondo (SCALED STO-3G)

1,2,4-6 dzp (DZP BASIS FOR ATOMS 1,2,4,5,6)

"c" tz (TZ BASIS FOR ALL CARBON ATOMS)

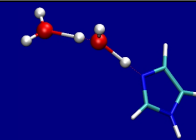
ANY DISPLAY COMMAND dis MAY BE ENTERED OR YOU MAY

HIT >return< TO QUIT (GOING BACK TO MAIN MENU)

all def2-SVP

*

TURBOMOLE single point for QM/MM (3/5)



CHOOSE COMMAND

infsao : OUTPUT SAO INFORMATION

atb : Switch for writing MOs in ASCII or binary format

eht : PROVIDE MOS && OCCUPATION NUMBERS FROM EXTENDED HUECKEL GUESS

use : SUPPLY MO INFORMATION USING DATA FROM

man : MANUAL SPECIFICATION OF OCCUPATION NUMBERS

hcore : HAMILTON CORE GUESS FOR MOS

flip : FLIP SPIN OF A SELECTED ATOM

& : MOVE BACK TO THE ATOMIC ATTRIBUTES MENU

THE COMMANDS use OR eht OR * OR q(uit) TERMINATE THIS MENU !!!

FOR EXPLANATIONS APPEND A QUESTION MARK (?) TO ANY COMMAND

eht

ENTER THE MOLECULAR CHARGE (DEFAULT=0)

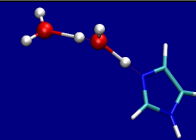
0

! put in here the total charge of you qm region

DO YOU ACCEPT THIS OCCUPATION ? DEFAULT=y

hit enter

TURBOMOLE single point for QM/MM (4/5)



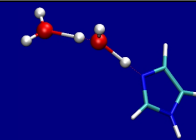
```
GENERAL MENU : SELECT YOUR TOPIC
scf : SELECT NON-DEFAULT SCF PARAMETER
mp2 : OPTIONS AND DATA GROUPS FOR rimp2 and mpgrad
...
```

```
dft
STATUS OF DFT_OPTIONS:
DFT is NOT used
functional b-p
gridsize m3
ENTER DFT-OPTION TO BE MODIFIED
func : TO CHANGE TYPE OF FUNCTIONAL
grid : TO CHANGE GRIDSIZE
on: TO SWITCH ON DFT
Just , q or '*' terminate this menu.
on
func b3-lyp
*

dsp
on
*

terminate define
*
```

TURBOMOLE single point for QM/MM (5/5)



in control file change:

```
$scfiterlimit 500
```

```
$thize 0.10000000E+99
```

```
$scfdamp start=1.700 step=0.050 min=0.050
```

remove line: `$scfdump`

(can also be done with "kdg")

run a scf iteration

```
dscf > dscf.out
```

in control file \$drvopt section add:

```
point charges
```

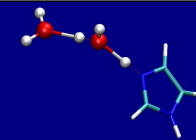
at the end of the control file add:

```
$point_charges file=point_charges
```

```
$point_charge_gradients file=pc_gradient
```

this will calculate a QM force on MM atoms, add MM charges around QM system, and calculate the gradients from MM on QM

Perform QM/MM single point energy evaluation



!stop comment this out

```
scalar mass show select type qq* show end
scalar mass set 1.008000 select type qq* show end
scalar mass show select type qq* show end
```

scale the mass of
ink atoms to 1.008

NBOND –

```
atom   cdiel  eps 1.0 –           ! Electrostatics
vatom  vswitch      –           ! VdW
elec   switch      –           ! Elec
ctonnb 100.0 ctofnb 120.0 cutnb 140.0
```

define cutoffs for
electrostatics and vdw
no PME yet for DFT/MM

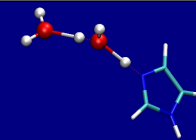
```
qturbo remove sele qm end
```

call turbomole
remove all MM contributions
for QM definition

ENERGY

evaluate QM/MM energy

Perform QM/MM single point energy evaluation



```
open write unit 41 file name "optimize-final.dcd"
```

open trajectory file

```
mini abnr nstep 500 nprint 1 tolg 0.0006 -  
tole 0.0006 IUNCrd 41 NSAVC 1
```

```
write coor pdb unit 10  
* minimized  
*
```

perform geometry optimization with
Adopted Basis Newton-Raphson for 500 se
or until tolerance in energy/gradient is
less than 0.0006 (kcal/mol)

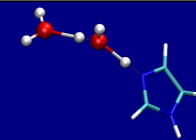
```
open write card unit 10 name "minimized.crd"  
write coor unit 10 card  
* minimized  
*
```

```
open write unit 10 card name "minimized.psf"  
write psf unit 10 card  
* minimized  
*
```

write coordinates and psf file

```
stop
```

Perform QM/MM MD

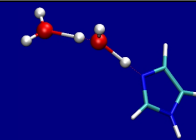


```
OPEN UNIT 44 write   FORMatted NAME "charmm_output/dynamics.res"
OPEN UNIT 42 write UNFORMatted NAME "charmm_output/dynamics.dcd"
open write unit 31 card name "charmm_output/dynamics.dat"
open unit 20 write uniform name "charmm_output/velocities.trj"
```

```
set STEP 500
SCALAR FBETA SET 8.0 select all end
```

```
dynamics lang leap start -
  timestep 0.001      - ! timestep in picoseconds
  nstep @STEP         - ! number of steps and energy evaluations
  nprint 1            - ! step frequency for writing in kunit and pr energy on unit 6
  iunwri 44           - ! unit to write restart file
  iunrea -1           - ! unit to read restart file
  iuncrd 42           - ! unit to write coordinates (unformatted)
  iunvel 20           - ! unit to write velocities out to
  kunit 31            - ! unit to write energy and temp inforamtion out to file
  iprfrq 20          - ! frequency for avg and rms energy
  firstt 310.         - ! initial T for velocity assignments
  tstruc 310.         - ! initial T for velocity assignments
  finalt 310.         - ! final T
  tbath 310.          - ! temperature of heat bath
  iasors 1            - ! asgn (NOT scale) veldur heat/equil (.ne. 0)
```

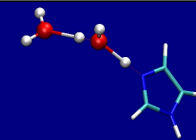
restarting QM/MM MD (1/2)



```
OPEN UNIT 11 read    FORMatted NAME "charmm_output/dynamics.res"  
OPEN UNIT 44 write   FORMatted NAME "charmm_output/dynamics2.res"  
OPEN UNIT 42 write UNFORMatted NAME "charmm_output/dynamics2.dcd"  
open write unit 31 card name "charmm_output/dynamics2.dat"  
open unit 20 write uniform name "charmm_output/dynamics2.trj"
```

```
set STEP 500  
SCALAR FBETA SET 8.0 select all end
```

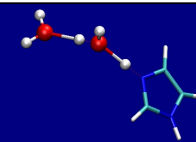
restarting QM/MM MD (2/2)



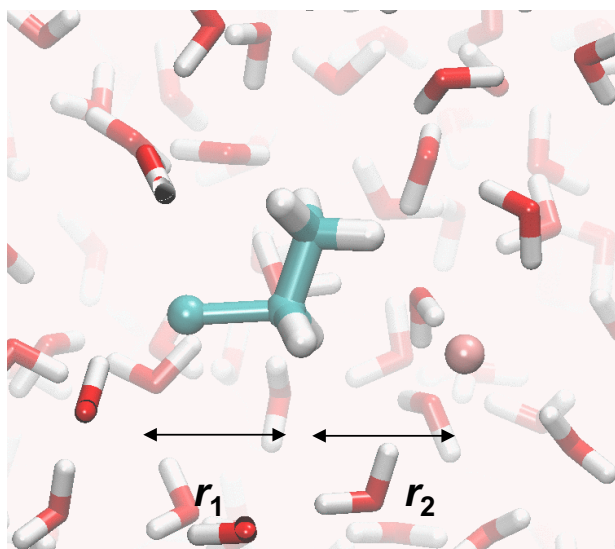
```
dynamics lang leap RESTRT -
  timestep 0.001      - ! timestep in picoseconds
  nstep @STEP         - ! number of steps and energy evaluations
  nprint 1            - ! step frequency for writing in kunit and pr energy on unit 6
  iunwri 44           - ! unit to write restart file
  iunrea 11           - ! unit to read restart file
  iuncrd 42           - ! unit to write coordinates (unformatted)
  iunvel 20           - ! unit to write velocities out to
  kunit 31            - ! unit to write energy and temp inforamtion out to file
  iprfrq 20          - ! frequency for avg and rms energy
  firstt 310.        - ! initial T for velocity assignments
  tstruc 310.        - ! initial T for velocity assignments
  finalt 310.        - ! final T
  tbath 310.         - ! temperature of heat bath
  iasors 1           - ! asgn (NOT scale) veldur heat/equil (.ne. 0 = asgn; .eq. 0 = scale)
  iasvel 1           - ! use gaussian distrib of vel (.gt. 0 =gaussian ; .lt. 0 = uniform)
  nsavc 1            - ! freq for writing coordinates
  nsavv 1            - ! freq for writing velocities
  isvfrq 100         - ! frequency for writing restart files
  ihtfrq 0           - ! frequency for heating steps during the dynamics
  iegfrq 0           - ! frequency for scaling velocities during heating
  inbfrq 1           - ! lists updated when necessary (heuristic test)
  ihbfrq 0           - ! frequency for updating hydrogen bond list
  ntrfrq 0           - ! step freq for stopping rotation and translation
  echeck 100000000.0 - ! max variation of energy step-to-step
  icheckw 0          - ! check to see if avg. temp. lies w/temp range (0 = do not check)
```

```
OPEN UNIT 1 WRITE CARD NAME "charmm_output/dynamics2.pdb"
WRITE COOR PDB UNIT 1
```

Example 2: Free energy surface of chemical reaction in solution



- QM/MM free energy simulation
 $\text{Cl}^- + \text{CH}_3\text{CH}_2\text{-Br} \rightarrow \text{Cl-CH}_3\text{CH}_2 + \text{Br}^-$



reaction coordinate:

$$\mathbf{R} = \mathbf{r}_1 - \mathbf{r}_2$$

employ bias on the reaction coordinate:

$$V_b = \frac{1}{2} k (\mathbf{R} - r_0)^2$$

run 1 $\rightarrow r_0 = -2.0$
(reactant: Cl-C is short, C...Br is long)

...

run 15 $\rightarrow r_0 = +1.7$
(product: Cl...C is long, C-Br is short)

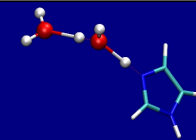


compute unbiased
prob. distr. $p(\mathbf{R})$
based on biased
data $p_b(\mathbf{R})$ and V_b



unbiased potential of
mean force for $\mathbf{R} \rightarrow G(\mathbf{R})$

How to set this up in practice?



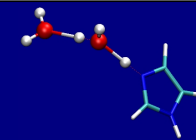
- Introduce restraints using the RESD module in CHARMM

```
SET ATOM1 PROA 1 CL  
SET ATOM2 PROA 1 C1  
SET ATOM3 PROB 1 BR
```

```
! 1.85-3.49 = -1.64  
SET V -1.64
```

```
SKIP NONE  
RESDistance RESET KVAL 100.0 RVAL @v -  
1.0 @atom1 @atom2 -1.0 @atom2 @atom3
```

How to set this up in practice?



- start by a short minimization that relaxes the restrained coordinate (as in example 1)

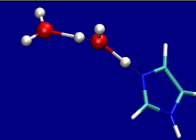
```
mini abnr nstep 10 nprint 1 tolg 0.0001 tole 0.0001 IUNCrd 41 NSAVC 1
```

```
open write card unit 10 name "mini.pdb"  
write coor pdb unit 10  
* minimized  
*
```

```
open write card unit 10 name "mini.crd"  
write coor unit 10 card  
* minimized  
*
```

```
open write unit 10 card name "mini.psf"  
write psf unit 10 card  
* minimized  
*
```

How to set this up in practice?



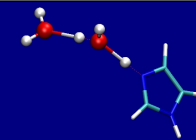
■ ... then run restrained dynamics

```
open write unit 42 file name "./trajectory.dcd"  
open unit 44 write card name "./dyn.res"  
open write unit 31 card name "./production.dat"  
open unit 20 write uniform name "./velocities3.trj"
```

```
! for 10000 x 1 fs steps  
set STEP 10000
```

```
! coupling for langevin dynamics  
SCALAR FBETA SET 8.0 select all end
```

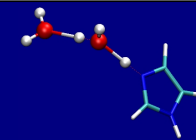
How to set this up in practice?



■ ... then run restrained dynamics

```
dynamics lang leap start -
  timestep 0.001          - ! timestep in picoseconds
  nstep @STEP             - ! number of steps and energy evaluations
  nprint 1                - ! step frequency for writing in kunit and pr energy on unit 6
  iunwri 44               - ! unit to write restart file
  iunrea -1               - ! unit to read restart file
  iuncrd 42               - ! unit to write coordinates (unformatted)
  iunvel 20               - ! unit to write velocities out to
  kunit 31                - ! unit to write energy and temp information out to file
  iprfrq 20               - ! frequency for avg and rms energy
  firstt 298.             - ! initial T for velocity assignments
  tstruc 298.             - ! initial T for velocity assignments
  finalt 298.             - ! final T
  tbath 298.              - ! temperature of heat bath
  iasors 1                - ! asgn (NOT scale) veldur heat/equil (.ne. 0 = asgn; .eq. 0 = scale)
  iasvel 1                - ! use gaussian distrib of vel (.gt. 0 =gaussian ; .lt. 0 = uniform)
  nsavc 1                 - ! freq for writing coordinates
  nsavv 1                 - ! freq for writing velocities
  isvfrq 100              - ! frequency for writing restart files
  ihtfrq 0                - ! frequency for heating steps during the dynamics
  iegfrq 0                - ! frequency for scaling velocities during heating
  inbfrq 1                - ! lists updated when necessary (heuristic test)
  ihbfrq 0                - ! frequency for updating hydrogen bond list
  ntrfrq 0                - ! step freq for stopping rotation and translation
  echeck 100000000.0      - ! max variation of energy step-to-step
  ichecw 0                - ! check to see if avg. temp. lies w/temp range (0 = do not check)
```

How to set this up in practice?



- ... and write final coordinates:

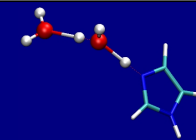
```
open write card unit 10 name "dynamics_all.pdb"
write coor pdb unit 10
* dynamics
*
```

```
open write card unit 10 name "dynamics_all.crd"
write coor unit 10 card
* dynamics
*
```

```
open write unit 10 card name "dynamics_all.psf"
write psf unit 10 card
* dynamics
*
```

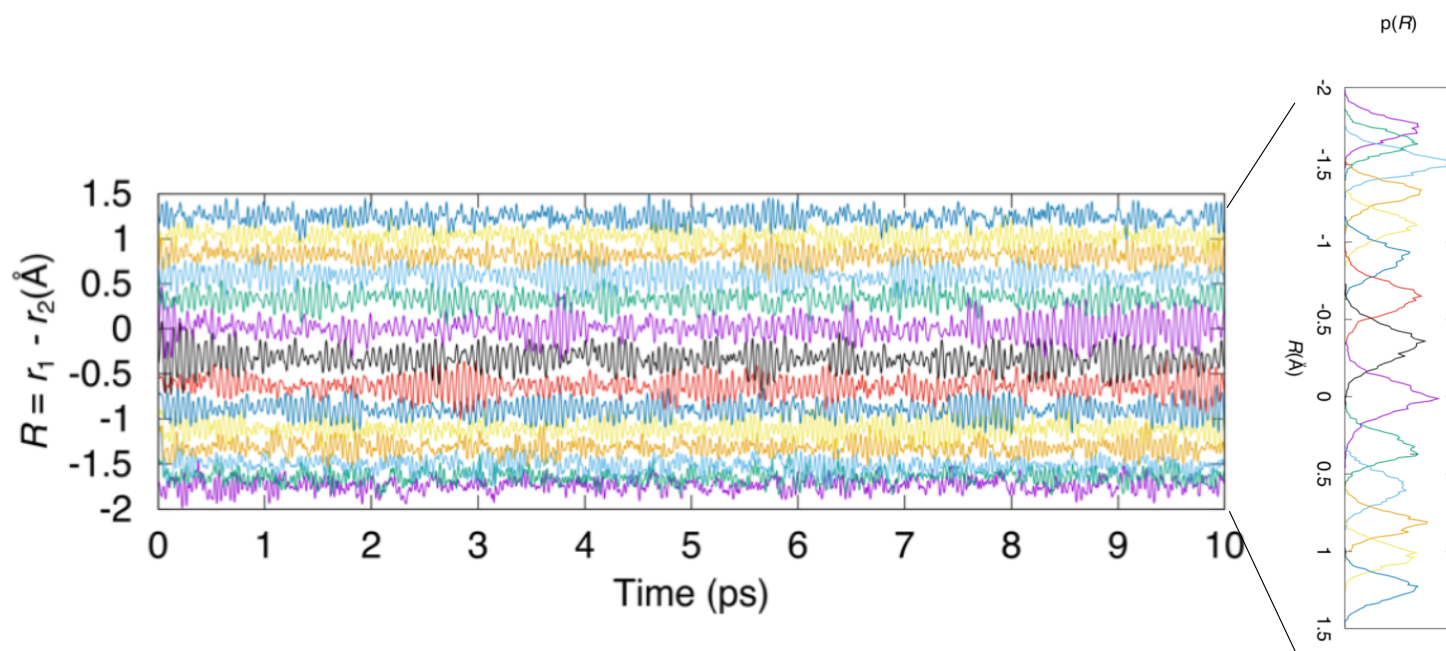
```
stop
```

How to set this up in practice?

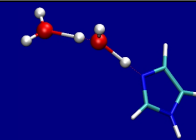


- extract distances from restrained runs

us_0.dat	us_11.dat	us_13.dat	us_15.dat	us_2.dat	us_4.dat	us_6.dat	us_8.dat
us_10.dat	us_12.dat	us_14.dat	us_1.dat	us_3.dat	us_5.dat	us_7.dat	us_9.dat



How to set this up in practice?



■ wham together

<http://membrane.urmc.rochester.edu/content/wham>

```
$ ./wham
```

```
Command line: wham [P|Ppi|Pval] hist_min hist_max num_bins tol temperature numpad  
metadatafile freefile [num_MC_trials randSeed]
```

```
./wham -2 1.5 16 0.000001 310 0 metadata.dat freefile
```

leave out P if not periodic

■ plot your data

```
gnuplot  
p 'freefile' u 1:2
```

Analyze your PMF

